

The CYPome of the model xenobiotic-biotransforming fungus

Cunninghamella elegans

William Palmer-Brown,¹ Raúl Miranda-CasoLuengo,¹ Kenneth H. Wolfe,² Kevin P. Byrne² and Cormac D. Murphy^{1*}

1. UCD School of Biomolecular and Biomedical Science, University College Dublin, Belfield, Dublin 4, Ireland

2. UCD School of Medicine, Conway Institute, University College Dublin, Belfield, Dublin 4, Ireland

Supplementary Information

Table S1. Primers used to amplify CYP genes from *C. elegans* gDNA and cDNA.

Primers	Nucleotide sequence 5' to 3'	T _m (°C)	CYP
g6049 F	TGGCACGAACAACCTTGGTAA	53.2	5206U1
g6049 R	CCAGCAGCAACTAAATCAGC	55.3	
g5706 F	GCTGATTTAGTTGCTGCTGGT	57.9	5208A2
g5706 R	CTAAACCCCGACCAATGGTA	55.9	
g5705 F	GCTGATTTAGTTGCTGCTGGT	55.3	5206D1
g5705 R	GGGAAAACTCTGCCATTTG	55.3	
g4598 F	TGGCAGAGTTTTTCCCTTTT	55.3	5205A8
g4598 R	TGAATCCCAACACAAACTCG	57.3	
g6826 F	GACTTGCTGTGCAACTTCAA	57.3	509Q1
g6826 R	TGGTATCTACACCAGCAGCAA	57.3	
g4063 F	GCTGACTTAATTGCTGCTGGT	53.5	5210C1
g4063 R	TGGTAATATAAGCTGCCATTC	55.3	
g3940 F	TGCACCACGTGCTATTAAGG	57.3	5313E1
g3940 R	ACGGGGATGAATGGTCATAA	57.3	
g2620 F	ATGAATGACTTTAATATTTACAATAAATTGGAAC	63.9	5313D1
g2620 R	TTAAAATTTTCTGGGCTTAACTTCATA	63.8	

Table S2. Primers used in RT-qPCR.

Primers	Nucleotide sequence 5' to 3'	T _m (°C)	GC-content (%)	T _a (°C)
CYP F*	GGTATGAACTTTAGTTTAGAACAA	55.9	25.0	58
CYP R*	CGGATTTTCAAATCAATTGGTTTAGGT	58.9	33.3	58
CPR F*	ATTACTGACGAATCCCCTGT	55.3	45	56
CPR R*	CGTTCACCAATACGTTTAGC	55.3	45	56
SarA F	AACCTACTCTTCATCCTACCTCTGAA	61.6	42.3	60
SarA R	GCATCAACTAAGAAAACATAACCAGA	58.5	34.6	60
UbcB F	ATGAGAGATTTCAAGAGACTCCAAA	58.1	36	60
UbcB R	ACAAATTTAACTGATGGTGGTTTGT	56.4	32	60
q-3940 F	ATTTGTTCTAAAGGCACTAAGGTT	57.3	50	60
q-3940 R	TGAAATTCATTAAATCATTGGGACT	57.3	50	60
q-2620 F	AATTCTACTGGATTTGCTATGATTG	53.5	30.4	60
q-2620 R	TTCATTGATGACAGCATTAAAGATAA	55.3	45	60

CYP and CPR primers were specific for CYP509A1 and a cytochrome P450 reductase, respectively.

Table S3. Conditions for RT-qPCR reactions

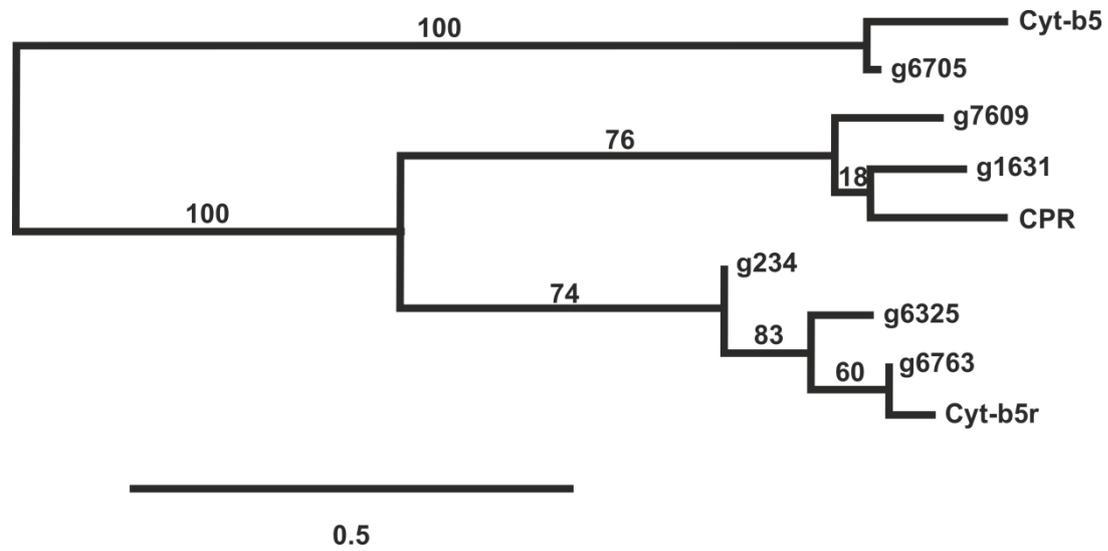
Step	Temperature (°C)	Time
Pre-incubation	95 °C	5 min
45 cycles	95 °C	15 s
	~ °C	20 s
	72 °C	10 s
Melting curve	50 to 95 °C	(temperature transition 0.2 °C /s)
Cooling	40 °C	10 s

Table S4. PCR efficiency of amplified targets*

Target	Slope	Y-intercept	E	Error	R2
<i>cyp5313E1</i>	-3.5196	28.12	1.87	0.72	0.94
<i>cyp5313D1</i>	-3.5354	26.31	1.92	0.27	0.99
<i>cyp509A1</i>	-3.2309	29.58	2.04	0.08	1
<i>ubcB</i>	-3.5794	24.5	1.9	0.2	1
<i>sarA</i>	-3.7986	24.62	1.83	0.34	0.99
<i>cpr</i>	-4.2994	25.45	1.71	0.32	0.99

*PCR efficiencies were measured from calibration curves made of serial dilutions of cDNA 1, 1/4, 1/16 and 1/64 measured in duplicate.

Figure S1. Phylogenetic tree of redox partner proteins from *C. elegans* and known CYP partner proteins used for homology searches. Branch bootstrap values are presented as a percentage.



G1631	QKLNPSVANPNHPPYAIEGPHQAYFQKNGDTIITKLPVHIRQSTFRLPTDASTPVMVGP	574
G7609	SIVHQVDDGRSYPHYDIAGPRQQLFESSL--SHAKIPIHVRRSQFKLPRNSKVPVIMVGP	586
G4301	MQNNDSDVIQGLPKYDLAGPRKAFNLNGQG--QSHKLP IHIRRSQFKLPRNTSCPVIMIGP	563
GAN10969.1	ESINGIEPDASFPHYSITGPRNCLYDTEA--KAARVPVHVRRSQFKLPRNPTVPIIMVGP	568
RCH89036.1	EAINGLEPDTASPRYYLGGPRDGLFDVQA--KQARVPVHVRRSQFKLPRNPTVPIIMVGP	565
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G1631	GTGVAPFRGFVRERVYQKQVEKQQVGTLLFFGCRRSNEDYLYADEWPELNFQLDNESRI	634
G7609	GTGVAPFRGFVRERALLKQK-GQEVGPTVLFFGCRHSEKDFIYKDEWPELNFNTLGEESQL	645
G4301	GTGVAPFRGFVRERERLQKKE-GKSVGPTILFFGNRHSQHDFLYSDEWPELNFNTLGEDSKL	622
GAN10969.1	GTGVAPFRGFVRERERLQKKE-GKPIGPTILYFGCRNSAEDFLYEEEWPELNFETLGESSRI	627
RCH89036.1	GTGVAPFRGFVRERERLQKKE-GKPVGTTVLFFGCRHPEQDFLYADEWPELNFETLGGDSRL	624
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G1631	INAFSRETDKKVYVQHRVRENGEEVWDLLANQGGYLYVCGDAKRMAKDITQTILDLAKHF	694
G7609	ITAFSRETQKVVYVQHRLEKEYGQQMWDYI-QQGAYIYVCGDAKNMAHDVNQAFIQFAQEF	704
G4301	ITAFSRESEHKVYVQHRLEENGKEIWQLL-EKGAYIYVCGDARNMARDVNQTFVNLAMEY	681
GAN10969.1	ITAFSRETAQKVYVQHRLEMENGPENWDLLEKGAIVYVCGDAKVMARDVNQTFVRFQQF	686
RCH89036.1	ITAFSRETDKKVYVQHRLLHEHGQAMWNLLEKGAIVYVCGDAKVMARDVNQTFVQFAKDL	683
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G1631	GKLDDEAALAFVQGLRKAGRYQEDVWA	721
G7609	GGRDEVKANAFFKQLRNTGRYQEDVWS	731
G4301	GEKTEQKALDYVKSLRNTGRYQEDVWS	708
GAN10969.1	GGHDEEKAQDYVKNLRNTGRYQEDVWS	713
RCH89036.1	GGLDEAEAQDYVKNLRNTGRYQEDVWA	710
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Figure S3. Amplification of *cyp5313D1* from cDNA of *C. elegans* DSM1908. The expected amplicon size was 1,539 bp.

